

Rec. Nat. Prod. 6:3 (2012) 301-305

records of natural products

Potent Insecticidal Secondary Metabolites from the Medicinal Plant Acanthus montanus

Elham Amin,^{1, 5} Mohamed M. Radwan,^{3,5} Seham S. El-Hawary,² Magda M. Fathy,² Rabab Mohammed,¹ James J. Becnel⁴ and Ikhlas Khan^{5*}

¹Faculty of Pharmacy, Beni Suef University, Beni Suef, Egypt ²Faculty of Pharmacy, Cairo University, Cairo, Egypt ³Faculty of Pharmacy, Alexandria University, Alexandria, Egypt. ⁴USDA, ARS, Center for Medical, Agricultural, and Veterinary Entomology Gainesville, FL 32608

USA.

^{5*}National Center for Natural Products Research, University of Mississippi, University, MS 38677,

USA.

(Received March 28, 2011; Revised October 7, 2011; Accepted October 10, 2011)

Abstract: Acanthus montanus (Nees) T. Anders. (Family: Acanthaceae) is a small shrub with sparse branches and soft stems, widespread in Africa, the Balkans, Romania, Greece and Eastern Mediterranean. Documented evidence showed that the leaves of the plant possess spasmolytic, analgesic, anti-inflammatory and antipyretic activities. In our ongoing research project; aimed at identifying new natural compounds with insecticidal activity, the alcohol extract of the aerial parts of *A. montanus* exhibited a significant activity against adult *Aedes aegypti*. Phytochemical study of the plant has resulted in isolation of nine compounds, eight of which exhibit variable degrees of insecticidal activity. β -sitosterol-3-O- β –D-glucoside (1) exhibited potent mosquitocidal activity (100% mortality) against adult *Aedes aegypti* at 1.25 µg/mg concentration, followed by palmitic acid (2) (90%), linaroside (3) (80%), and acetoside (9) (70%) respectively. It is noteworthy that this is the first report of insecticidal activity of β -sitosterol-3-O- β –D-glucoside and acetoside.

Keywords: Acanthaceae, Acanthus montanus, adult Aedes aegypti, β -sitosterol glucoside, palmitic acid, protochatecuic acid, shikimic acid.

1. Plant Source

Acanthus montanus (Nees) T. Anders. (Family: Acanthaceae) is a small shrub with sparse branches and soft stems, widespread in Africa, the Balkans, Romania, Greece and the Eastern Mediterranean [1]. Aerial parts of the plant were collected from Cameroon in March 2003. Botanical identification was performed by Trish Flaster, the executive director of Botanical Liaisons, a voucher specimen, number ACM-2326, was deposited at the repository of National Center for Natural Products Research, School of Pharmacy, University of Mississippi, USA.

Medicinal plant Acanthus montanus

2. Previous Studies

Previous biological investigations have shown that; the leaves of *A. montanus* display spasmolytic, analgesic, anti-inflammatory and antipyretic activities [2-4]. In Cameroon the plant is used traditionally to treat various ailments namely; cough, carious teeth, pharyngitis, dysmenorrhoea, gastritis, false labour, epilepsy, and intestinal helminthiasis, in other regions of Africa it is used to alleviate uretheral discharge, chest pain, emesis, constipation, rheumatic pains, and syphilis [5-6].

Many species of genus Acanthus have been investigated and shown to contain several classes of secondary metabolites, particularly benzoxazinoides, phenylethanoides, lignans, flavonoides, megastigmanes, fatty acids and aliphatic alcohol glycosides [7-20]. Previous phytochemical studies of *A. montanus* reported the isolation of triterpenoid glycosides [21-22], phenyl ethanoides, bezoxazinoides and fatty alcohols [23]. However, no relevant literature, substantiating the evaluation of the insecticidal activity of the plant, has been reported.

3. Present Study

In our ongoing research project, aimed at identifying natural plant constituents with pesticidal activity [24], extracts of several plants were evaluated. The alcohol extract of the aerial parts of *A. montanus* displayed a significant activity against adult female *A. aegypti*, so it was chosen for the study. Aerial parts of the plant 400 g were exhaustively extracted with 80% MeOH, and the dried extract was subjected to several chromatographic techniques to yield nine compounds (1-9). The isolated compounds were tested for insecticidal activity against *A. aegypti*, using the adult assay protocol established by Pridgeon *et al.*, [25-26]. Detailed phytochemical and pharmacological procedures are provided as supporting information.

3.1. Identification of the isolated compounds.

On the basis of spectral data obtained from HR-ESI-MS and extensive NMR experiments (¹H, ¹³C, APT, DEPT-135, COSY, HMQC, HMBC and NOESY) and by comparison with spectral values reported in the literature [27-33], the compounds obtained from *A. montanus* were identified as; β -sitosterol-3-O- β –D-glucoside (1), palmitic acid (2), linaroside (3), homoplantagenin (4), 5, 7, 3'-trihydroxy-6,4' -dimethoxy flavone-7-O-glucoside(5), shikimic acid (6), protochatecuic acid (7), blepharin (8), and acetoside (9). It is noteworthy that this is the first report of isolation of (1—7) from *A. montanus*. The isolated compounds belong to different chemical classes; fatty acids, flavonoides, sterol glucoside, benzoxazinoide, phenolic acid, and phenyl ethanoid. Accordingly, the present work, confirms the typical profile of secondary metabolites found in the genus Acanthus. Moreover, it reports the first isolation of the two acids; protochatecuic and shikimic, from family Acanthaceae. Hence, adding another class of secondary metabolites not previously reported from this family. The occurrence of such metabolites provides a chemotaxonomic tool for further studies of this genus.

3.2. Insecticidal activity testing results

Eight of the isolated compounds were tested for activity against female adults of *A. aegypti*, at two concentrations; 1.25 µg/mg and 0.63 µg/mg. β -sitosterol glucoside (1) and palmitic acid (2), were the most active showing 100% and 90% mortality, respectively, at 1.25 µg/mg concentration, followed by linaroside (3) 80%, acetoside (9)70%, protochatecuic acid (7) 40%, and homoplantagenin (3) 30%. At concentration 0.63 µg/mg, compounds (1), (2), (3), and (6) were active showing 90%, 80%, 70%, and 10% adulticidal activity, respectively.

Acetone and Permethrin were used as negative and positive controls causing 0% and 100% mortality, respectively, at the tested concentrations. Permethrin $LD_{50} = 4.9 \times 10^{-5}$ [34].

The larvicidal activity of palmitic acid, against *Culex quinquefasciatus*, *Anopheles stephensi* and *A. aegypti*, was previously reported by Abdul-Rahman *et al.* 2000 [35], however, this is the first report of the adulticidal activity of these compounds.



Figure 1: Compounds isolated from Acanthus montanus



Figure 2: Mosquitocidal activity of isolated compounds

Acknowledgments

Elham Amin thanks the Egyptian Government for the fellowship through The Ministry of Higher Education and Scientific Research. This study was also supported by a grant from Deployed War-Fighter Protection Research Program (DWFP), the U.S. Department of Defense through the Armed Forces Pest Management Board (AFPMB) and the USDA ARS NPURU.

Supporting Information

Supporting information accompanies this paper on http://www.acgpubs.org/RNP

References

- [1] C. O. Okoli, P. A. Akah, N. J. Onuoha, T. C. Okoye, A. C. Nwoye and C. S. Nworu (2008). Acanthus montanus: An experimental evaluation of the antimicrobial, anti-inflammatory and immunological properties of a traditional remedy for furuncles, BMC Complement Altern. Med. 8, 27.
- [2] O. O. Adeyemi, S. O. Okpo and C. C. Young-Nwafor (1999). The relaxant activity of the methanolic extract of *Acanthus montanus* on intestinal smooth muscles, *J. Ethnopharmacol.* **68**,169-173.
- [3] O. O. Adeyemi, S. O. Okpo and O. Okpaka (2004). The analgesic effect of the methanolic extract of *Acanthus montanus*, *J. Ethnopharmacol.* **90**, 45-48.
- [4] E. A. Asongalem, H. S. Foyet, S. Ekobo, T. Dimo and P. Kamtchouing (2004). Anti-inflammatory, lack of central analgesia and antipyretic properties of *Acanthus montanus* (Ness) T. Anderson, *J. Ethnopharmacol.* 95, 63-68.
- [5] J. E. Adjanohoun, N. Aboubakar, K. Dramane, M. E. Ebot and J. A. Ekpere (1996). OAU/STRC Traditional Medicine and Pharmacopoeia: Contribution to Ethnobotanical and Floristic studies in Cameroon. CNPMS, Porto Novo, Benin, pp. 360-420.
- [6] H. M. Burkill (1985). Useful Plants of West Africa, 1. Royal Botanic Garden, pp. 458-459.
- [7] M. E. Amer, M. I. Abou-Shoer, M. S. Abdel-Kader, A. M. S. El-Shaibany and N. A. Abdel-Salam (2004). Alkaloids and Flavone Acyl Glycosides from *Acanthus arboreus, J. Braz. Chem. Soc.* **15**, 262-266.
- [8] S. Goswami, B. Chatterjee and M. Mallik (2004). Proof of presence of a complete homologous series of odd and even numbered fifteen saturated fatty acids in *Acanthus ilicifolius*. L (Acanthaceae), J. Indian Chem. Soc. 81, 696-706.
- [9] C. Huo, D. An, B. Wang, Y. Zhao and W. Lin (2005). NMR spectral assignments of a new benzoxazolinone glucoside from *Acanthus ilicifolius*, *Magn. Reson. Chem.* **43**, 343-345.
- [10] C. Huo, B. Wang, W. Lin and Y. Zhao (2005). Benzoxazinones from Acanthus ilicifolius, Biochem. Syst. Ecol. 33, 643-645.
- [11] T. Kanchanapoom, M. S. Kamel, R. Kasai, K. Yamasaki, C. Picheansoonthon and Y. Hiraga (2001). Lignan glycosides from *Acanthus ilicifolius*. *Phytochemistry* **56**, 369-372.
- [12] T. Kanchanapoom, M. S. Kamel, R. Kasai, C. Picheansoonthon, Y. Hiraga and K. Yamasaki (2001). Benzoxazinoid glucosides from *Acanthus ilicifolius, Phytochemistry* **58**, 637-640.

- [13] T. Kanchanapoom, R. Kasai, C. Picheansoonthon and K. Yamasaki (2001). Megastigmane, aliphatic alcohols and benzoxazinoid glycosides from *Acanthus ebracteatus*, *Phytochemistry* **58**, 811-817.
- [14] T. Kanchanapoom, P. Noiarsa, H. Otsuka and S. Ruchirawat (2006). Chemical constituents from *Acanthus volubilis* (Wall.), *Biochem. Syst. Ecol.* **34**, 442-445.
- [15] U. Kokpol, V. Chittawong and D. H. Miles (1986). Chemical constituents of the roots of *Acantus ilicifolus*. *J. Nat. Prod.* **49**, 355-356.
- [16] K. Pratt, P. Kumar and W. S. Chilton (1995). Cyclic hydroxamic acids in dicotyledonous plants, *Biochem. Syst. Ecol.* 23, 781-785.
- [17] R. B. Wolf, G. F. Spencer and R. D. Plattiner (1985). Benzoxazolinone-2, 4-dihydroxy-1, 4-benzoxazin-3one and its glucosides from *Acanthus mollis* seeds inhibit velvet leaf germination and growth, *J. Nat. Prod.* 48,59-63.
- [18] J. Wu, S. Zhang, Q. Xiao, Q. Li, J. Huang, L. Long and L. Huang (2003). Phenyl ethanoid and aliphatic alcohol glycosides from *Acanthus ilicifolius, Phytochemistry* **63**, 491-495.
- [19] J. Wu, S. Zhang, J. Huang, Q. Xiao, Q. Li, L. Long and L. Huang (2003). New aliphatic alcohol and (2)-4coumaric acid glycosides from *Acanthus ilicifolius, Chem. Pharm. Bull.* 51, 1201-1203.
- [20] J. Wu, S. Zhang, Q. Xiao, Q. Li, J. Huang, L. Long and L. Huang (2003). Megastigmane and flavone glycosides from *Acanthus ilicifolius, Pharmazie* **58**, 363-364.
- [21] E. M. Anam (1997). Pentacyclic triterpenoids from *Acanthus montanus* (Acanthaceae), *Ind. J. Chem.* **36B**, 110-113.
- [22] E. M. Anam (1997). Novel triterpenoid glycoside and triterpenoid acid from the root extract of *Acanthus montanus* (Acanthaceae), *Ind. J. Chem.* **36B**, 901-904.
- [23] P. Noiarsa, S. Ruchirawat and T. Kanchanapoom (2010). Acanmontanoside, a new phenyl ethanoid diglycoside from *Acanthus montanus*. *Molecules*. **15**, 8967-8972.
- [24] E. Amin, S. S. El-Hawary, M. M. Fathy, R. Mohammed, A. Zulfiqar, N Tabanca, D. E. Wedge, J. Becnel and I. A. Khan (2010). Triterpenoidal saponins: bioactive secondary metabolites from *Zygophyllum coccineum*, *Planta Med.* doi: 10.1055/s 0030-1250463.
- [25] E. J. Gerberg, D. R. Barnard and R. A. Ward (1994). Manual for mosquito rearing and experimental techniques, *Am. Mosq. Control Assoc. Bull.* **5**.
- [26] J. W. Pridgeon, R. M. Pereira, J. J. Becnel, S. A. Allan, G. G. Clark and K. J. Linthicum (2008). Susceptibility of *Aedes aegypti*, *Culex quinquefasciatus* Say, and *Anopheles quadrimaculatus* Say to 19 pesticides with different modes of action, *J. Med. Entomol.* 45, 82-87.
- [27] E. A. Aboutabl, F. A. Hashem, A. A. Sleem and A. A. Maamoon (2008). Flavonoides, anti-inflammatory activity and cytotoxicity of *Macfadyena unguis*, *Afr. J. Trad. CAM.* **5**, 18-26.
- [28] L. C. Santos, C. M. Rodrigues, R. G. Coelho, M. Sannomiya and W. Vilegas (2005). Chemical profile of *Eriocaulon ligulatum* (Vell.) L.B. Smith (Eriocaulaceae), *Biochem. Syst. Ecol.* 33, 1159-1166.
- [29] L. Toth, I. Csordas, V. Papay and G. Y. Bujtas (1978). Flavonoids of Kickxia elatine (L.) Dum, *Pharmazie*, 33, 374-375.
- [30] L. B. Enrich, M. L. Scheuermann, A. Mahadjer, K. R. Matthias, C. F. Eller, S. Newman, M. Fujinaka and T. Poon (2008). *Liquidambar styraciflua*: a renewable source of shikimic acid. *Tetrahedron Letters*. 49, 2503-2505.
- [31] H. L. Zhang, A. Nagatsu, H. Okuyama, H. Mizukami and J. Sakakibara (1998). Sesquiterpene glycosides from cotton oil cake, *Phytochem.* **48**, 665-668.
- [32] L. F. Tietze, M. Beller, A. Terfort and A. Doelle (1991). First synthesis and structural determination of Blepharine and 1'-Epiblepharine, *Synthesis*. 1118-1120.
- [33] T. Tanaka, T. Ikeda, M. Kaku, X. H. Zhu, M. Okawa, K. Yokomizo, M. Uyeda and T. Nohara (2004). A new lignan glycoside and phenyl ethanoid glycosides from *Strobilanthes cusia* Bremek. *Chemical and pharmaceutical bulletin.* 52, 1242-1245
- [34] J. W. Pridgeon, J. J. Becnel, G. G. Clark and K. J. Linthicum (2009). A high throughput screening method to identify potential pesticides for mosquito control, *J. Med. Entomol.* **46**, 335-341.
- [35] A. AbdulRahman, G. Gopalakrishnan, B. S Ghouse, S. Arumugam and B. Himalayan (2000). Effect of *Feronia limonia* on mosquito larvae, *Fitoterapia*, **71**, 553.



© 2012 Reproduction is free for scientific studies